REVIEW ARTICLE



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Testosterone recovery therapy targeting dysfunctional Leydig cells

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Abstract

Reduced serum testosterone affects millions of men across the world and has been linked to several comorbidities, metabolic dysfunctions, and quality of life changes. The standard treatment for testosterone deficiency remains testosterone replacement therapy. However, limitations on its use and the risk of significant adverse effects make alternative therapeutics desirable. Studies on the mechanisms regulating and synthesizing testosterone formation in testicular Leydig cells demonstrate numerous endogenous targets that could increase testosterone biosynthesis, which could alleviate reduced testosterone effects. Testosterone biosynthesis is facilitated by a conglomerate of cytosolic and mitochondrial proteins that facilitate cholesterol translocation into the mitochondria, the rate-limiting step in steroidogenesis. An effective therapeutic approach would be required to increase endogenous testosterone formation by enhancing steroidogenesis in Leydig cells. Numerous ligands for steroidogenic proteins have been developed, which increase steroid hormone formation. However, off-target effects on neurosteroid and adrenal steroid formation may limit their clinical use. First-in-class biologics, such as voltage-dependent anion channel peptides and transplantation of induced human Leydig-like cells offer advances in the development of specific strategies that could be used to enhance endogenous steroid formation in hormone deficient patients.

KEYWORDS

aging, hypogonadism, infertility, testis, testosterone deficiency

1 | INTRODUCTION

Although some testosterone decline is normal in men of middle and advanced age, some men have significantly decreased testosterone levels known as hypogonadism. Hypogonadism is a condition characterized by severe testosterone deficiency and affects nearly 5 million men in the United States. While hypogonadism is most commonly associated with infertility, it has also been correlated with other

numerous conditions, such as cardiovascular disease, depression, fatigue, reduced bone mineral density, increased body fat, metabolic syndrome, and declining muscle mass. Hypogonadism can be separated into two categories: primary hypogonadism and secondary hypogonadism. Primary hypogonadal patients present depleted testosterone levels because of a suboptimal response to luteinizing hormone (LH) stimulation; whereas secondary hypogonadism is characterized by low LH levels or low gonadotropin releasing hormone (GnRH) levels,

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leading to insufficient steroid hormone biosynthesis.⁴ Moreover, primary hypogonadal patients display increased LH, suggesting that Leydig cell mechanisms are disrupted.⁵ The primary causes of secondary hypogonadism are associated with the pituitary or hypothalamus.⁴ These can be congenital, acquired, or caused by damage to gonadotrophs.⁴

Given testosterone's essential role in spermatogenesis, hypogonadal patients suffer from infertility. Furthermore, androgen metabolites levels, such as dihydrotestosterone (DHT) and 3α -androstenediol glucuronide, become imbalanced and cause alterations in secondary sex characteristics, including muscle mass, body mass index, and facial hair. Patients may present with fatigue and declining mood, given the ability of neurosteroids to act as positive or negative regulators of the gamma-aminobutyric acid receptor. There are also numerous congenital and acquired origins of hypogonadism that may manifest throughout the male lifespan. Therapeutic strategies for endogenous targets to treat hypogonadism from all origins are highly sought.

Testosterone replacement therapy (TRT)⁹ and aromatase inhibitors 10 have been used to elevate serum testosterone and alleviate symptoms of hypogonadism. TRT involves administering exogenous testosterone at appropriate intervals: either daily acting, intermediate acting (1-3 weeks), or long acting (2-6 months).9 However, this exogenous testosterone leads to hypothalamicpituitary-gonadal axis (HPG) imbalance and suppresses the release of gonadotropins. 11 This represses Leydig cell testosterone biosynthesis, a critical driver of spermatogenesis, and leads to reduced fertility. 9,11 Moreover, intermediate- and long-acting injections may produce serious adverse events including pulmonary microembolism, anaphylaxis, and polycythaemia, 9,12,13 and an increased risk of cardiovascular disease and stroke may exist in older men receiving TRT as indicated in recent studies, 14,15 resulting in the Food and Drug Administration and medical societies cautioning its use.³ Numerous alternatives to TRT have been considered. 16 The testosterone metabolite DHT is also used strategically to treat hypogonadism in some countries.¹⁷ DHT binds to androgen receptors with a greater affinity than testosterone and provides some relief from symptoms of hypogonadism. 18 The disadvantages of DHT are its price, increased hemoglobin, increased red blood cell count, and inferior clinical results than TRT.^{17,18} Aromatase inhibitors are also used to prevent aromatase from converting testosterone to estrogen, thereby, maintaining testosterone levels. 19 In clinical studies with aromatase inhibitor used for hypogonadal patients, LH levels, free testosterone, and sexual desire increased.²⁰ Moreover, aromatase inhibitors may be suitable for hypogonadal patients with increased estrogen levels. 18 However, concerns regarding the effect of aromatase inhibitors on bone minerals still remain after treatment with the inhibitor letrozole led to vertebrae deformities in 45% of adolescent males with delayed puberty.²¹ The selective estrogen receptor modulators clomiphene citrate and tamoxifen are also used off-label for the treatment of primary hypogonadism because of their ability to induce the release of GnRH by the hypothalamus and subsequently increase the production of the gonadotropins LH and follicle stimulating hormone by the anterior pitutary.16

2 | TESTOSTERONE REGULATION AND FORMATION

Testosterone biosynthesis predominantly occurs in testicular Leydig cells and is tightly regulated by the HPG axis, comprised of the hypothalamus, pituitary, and testes.²² In this system, the hypothalamus secretes GnRH, which reaches and stimulates the anterior pituitary gland to release LH. LH acts on the testicular Leydig cell LH receptor (LHR), a G protein-coupled receptor, and initiates a signaling cascade that mobilizes cholesterol and increases testosterone biosynthesis.²² LHR stimulation activates adenylate cyclase and increases cyclic adenosine monophosphate (cAMP) production and subsequent cAMP-dependent kinase activation.¹ Mechanistic targets inducing the production of endogenous testosterone in Leydig cells would be most desirable. Viable drug targets should have specificity, a sustainable response, and acceptable safety profiles.

The rate-limiting step in steroid hormone biosynthesis is cholesterol's translocation across the outer and inner mitochondrial membranes (OMM and IMM) into the mitochondria. Cholesterol's translocation into the IMM results in cholesterol side-chain cleavage by the cytochrome P450 CYP11A1, producing pregnenolone.²³ This translocation is mediated through a multiprotein scaffold termed as the Steroidogenic InteracTomE (SITE).²⁴ The SITE is comprised of cytosolic and mitochondrial proteins, of which numerous have become focal points in the search for endogenous targets that induce steroidogenesis. Cytosolic SITE proteins include the acyl-CoA-binding protein (ACBD1/DBI),²⁵⁻²⁷ ACBD3,²⁴ Sec23-interacting protein,²⁴ steroidogenic acute regulatory protein (STAR), 28-32 14-3-3 proteins, 33-35 and the cAMP-dependent protein kinase (PKA), which is composed of regulatory and catalytic subunits inducing STAR phosphorylation upon cAMP activation.³⁶ OMM SITE proteins include the translocator protein (TSPO),1 the voltage-dependent anion channel (VDAC1),1 and ATPase family AAA domain-containing protein 3A (ATAD3A),²⁴ while IMM SITE proteins include the cholesterol side-chain cleavage enzyme (CYP11A1), ferredoxin (FDX), and ferredoxin reductase (FDR).²⁴ The fine details of cholesterol's translocation across the mitochondrial membranes are not yet clear, but there are notable protein-protein interactions that have been elucidated (Figure 1).

VDAC1 and TSPO are the main anchors of the cytosolic proteins to mitochondrial contact sites. ^{24,37} ATAD3A bridges the mitochondrial membranes and is involved in contact site formation, mediating access of cholesterol to CYP11A1. ^{38,39} The adenine nucleotide translocase protein interacts strongly with VDAC1 to form a contact site complex between the OMM and IMM, which is involved for the trafficking of molecules across the mitochondrial membranes, ⁴⁰ but does not interact directly with the SITE complex as currently identified. ³⁸ In addition, the IMM optic atrophy 1 (OPA1) protein participates in the formation of contact sites and mitochondrial fusion between mitochondrial membranes, a process essential for steroidogenesis. ⁴¹ External response to hormonal stimulation initiates STAR targeting to the SITE complex at the OMM. ⁴² STAR anchors to the mitochondrial SITE scaffold at VDAC1, a solute-specific transporter to the IMM, ⁴⁰ and STAR becomes phosphorylated by PKA. ²³ PKA

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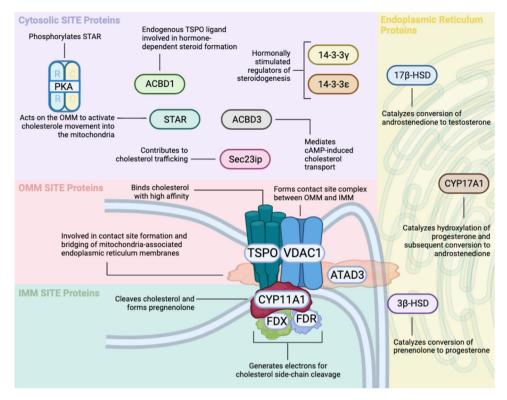


FIGURE 1 Steroidogenic InTeractomE (SITE) proteins of the Leydig cell. Cytosolic, outer mitochondrial membrane (OMM), inner mitochondrial membrane (IMM), and endoplasmic reticulum proteins interact to facilitate the transfer of cholesterol into the mitochondria and production of numerous steroid hormones, including testosterone in the endoplasmic reticulum. Abbreviations: 3β -HSD, 3β -hydroxysteroid dehydrogenase; 17β -HSD, 17β -hydroxysteroid dehydrogenase; ACBD1, acetyl coenzyme A-binding domain 1 or diazepam binding inhibitor; ACBD3, acetyl coenzyme A-binding domain 3; ATAD3A, ATPase family AAA domain-containing protein 3A; CYP11A1, cytochrome P450 11A1; CYP17A1, cytochrome 17A1; FDR, ferredoxin reductase; FDX, ferredoxin; PKA, cAMP-dependent protein kinase; PKA-R, regulatory subunit; PKA-C, catalytic subunit; Sec23ip, Sec23-interacting protein; STAR, steroidogenic acute regulatory protein; TSPO, translocator protein; VDAC1, voltage-dependent anion channel 1

is targeted to mitochondria by A-kinase anchoring proteins binding to the regulatory subunits to PKA, such as ACBD3,43 a protein that interacts with TSPO, and AKAP121.44 leading to effective translation and phosphorylation of STAR and conformational changes which would accelerate cholesterol translocation and optimize steroid formation.^{23,28} In response to these changes, TSPO is polymerized and cholesterol binding is enhanced⁴⁵ because of TSPO's high affinity for cholesterol. 38,46,47 TSPO contains five transmembrane domains with separate cholesterol and drug binding domains and is highly abundant in the OMM. 48 ACBD1/DBI is an endogenous ligand of TSPO, involved in hormone-dependent steroid formation.²⁴ The polymerization of TSPO strengthens the TSPO-VDAC1 interaction, enhancing cholesterol binding and transport. 36,38,49 SITE optimization enhances cholesterol translocation across the mitochondrial membranes to CYP11A1, where FDX and FDR regulate the electrons needed for side-chain cleavage by the enzyme. 24,38,48 14-3-3 γ and 14-3-3 ε are hormonally stimulated and act as negative regulators of steroidogenesis by delaying maximal steroid hormone formation.³³ Upon hormone stimulation, $14-3-3\gamma$ interacts with STAR, limiting its activity in cholesterol transport. 33 Similarly, stimulation also triggers 14-3-3ε binding to the VDAC1-TSPO complex and regulates cholesterol translocation into the mitochondria by reducing the rate of transport.³³ Other intracellular regulators of steroidogenesis include

signaling molecules (platelet-derived growth factor (PDGF), desert hedgehog (DHH), kinases mitogen-activated protein kinase (MAPK), protein kinase G (PKG), calcium/calmodulin-dependent protein kinase I (CAMKI), 5' AMP-activated protein kinase (AMPK), and transcription factors (nuclear receptor 4A1 also known as NUR77), myocyte enhancer factor 2 (MEF2), GATA binding protein 4 (GATA4).50,51 Moreover, numerous nuclear receptors and protein phosphorylation events are involved in steroidogenesis regulation. 52,53 Steroidogenesis is also regulated systemically by the HPG axis. 1 It is imperative that steroid hormone synthesis is precisely regulated, as insufficient or overproduction of steroids is detrimental.¹

3 | MECHANISMS OF LEYDIG CELL **DYSFUNCTION**

The physiopathology of numerous diseases related to impaired steroid hormone biosynthesis are mediated by compromised Leydig cell integrity. In aging, the integrity of Leydig cell-specific mechanisms mediating steroid hormone biosynthesis is compromised. Whereas gene mutations in key steroidogenic genes can lead to disease phenotypes or lethality, compromised Leydig cell integrity can be caused by several intracellular factors.

FIGURE 2 Off-target effects of therapeutic strategies. Numerous therapeutics that are used to treat testosterone deficiency have off-target effects on the hypothalamic pituitary gonadal axis, adrenal gland, and testicular Leydig cells. Abbreviations: GnRH, gonadotropin releasing hormone; TSPO, translocator protein; VDAC1, voltage-dependent anion channel 1

3.1 Reductions in steroidogenic enzymes

The steroidogenic machinery tightly regulates and maintains steroid hormone biosynthesis. Declining or aberrant expression of SITE proteins or other proteins involved in steroidogenesis can occur at the transport, import, or conversion steps. For example, STAR is constitutively expressed in Leydig cells, mediating cholesterol transport from intracellular stores to the mitochondria, and extracellular hormonal stimulation of Leydig cells increases STAR expression to upregulate cholesterol translocation. Mutations in *Star* lead to steroid hormone biosynthesis deficiency and the accumulation of lipids in testosterone-producing cells. Moreover, decreased *Star*/STAR expression with age reduces cholesterol translocation in aged Leydig cells. S8

Mutations to TSPO also alter the ability of steroidogenic cells to import cholesterol into the mitochondria. This results in increased lipid accumulation and disruption of steroid production and has implications for the hormone biosynthesis in the brain, adrenal glands, and testis. S9-61 TSPO's decline in aging Leydig cells showed that alterations in cholesterol import play a role in age-related testosterone decline. Other downstream steroidogenic enzymes that are decreased in aging include CYP11A1, HSD3B, CYP17A1, and HSD17B. 63

3.2 | Imbalanced antioxidant and reactive oxygen species production

Reactive oxygen species (ROS) are mostly produced by the mitochondria and can compromise the integrity of cellular machinery and structures.⁶⁴ Age-related oxidant/antioxidant imbalances are correlated with protein, lipid, and DNA damage, linking integrity of mitochondrial quality control to the development of age-related pathologies.⁶⁵ Oxidant/antioxidant imbalance may arise from increased oxidant production in Levdig cells, as mitochondrial superoxide production has been observed in aged rat Leydig cells.⁶⁶ While the generation of adenosine triphosphate (ATP) via the electron transport chain produces ROS ubiquitously in mammalian cells, Leydig cells also produce ROS through hormone biosynthesis via mitochondrial and smooth endoplasmic reticulum P450 reactions.^{67,68} Changes in biosynthesis can, thus, alter ROS production. Over time, ROS exposure damages mitochondria and compromises their function, leading to mitochondrial dysfunction.⁶⁹ When left uncleared, dysfunctional mitochondria produce excessive amounts of ROS which further damage cellular enzymes and structures. 65,70 Dysfunctional mitochondrial are normally eliminated via mitochondrial autophagy (mitophagy) and replaced by new mitochondria through mitochondrial biogenesis.⁷¹

TABLE 1 Treatment options available for testosterone deficiency and their use in other indications

| Therapy | Indications | Results | Concerns |
|----------------------------------|--|---|--|
| Testosterone replacement therapy | Hypogonadism Diabetes Osteoporosis Sexual desire | Testosterone ↑ Health ↑ Sexual function ↑ Bone mineral density ↑ Spermatogenesis ↓ Fertility ↓ Mood ↓ | Infertility, LH suppression, prostate cancer, fluctuating testosterone levels, lower hematocrit, skin irritation, development of male breast tissue, alterations in mood |
| Aromatase inhibitors | Hypogonadism (off-label) Breast cancer Gynecomastia Ovulation induction | Testosterone ↑ Bone mineral density ↓ | Hot flashes, weight gain, insomnia, venous thromboembolism, erectile dysfunction, breast pain |
| Gonadotropins | Hypogonadism Infertility (off-label) | Testosterone ↑ Spermatogenesis ↑ | Gynecomastia, erythrocytosis |
| Pulsatile GnRH therapy | Hypogonadism Fertility Prostate cancer Transgender therapy | Testosterone ↑ Spermatogenesis ↑ | Erythrocytosis, difficult, expensive |
| Tamoxifen/clomiphene citrate | Hypogonadism Dysmenorrhea Breast cancer Infertility Gynecomastia | Testosterone ↑ Spermatogenesis ↑ | Venous thromboembolism, vision, mood changes, weight gain, not effective in primary hypogonadism, off-label use |
| Dopamine agonists | Hypogonadism Psychopathological disorders Parkinson's disease | Testosterone ↑ Sexual function ↑ Semen quality ↑ Bone mineral density ↑ | Headaches, hypotension, nausea |
| Reduction of opioid use | Opioid-induced hypogonadism | Testosterone ↑ Sexual function ↑ | Few alternatives for treating chronic pain |
| Lifestyle changes | Health Wellbeing Hypogonadism Sexual function | Testosterone ↑ Sexual function ↑ Spermatogenesis ↑ Health ↑ Wellbeing ↑ | Time, motivational, and environmental barriers |
| TSPO ligands | Psychopathological disorders Hypogonadism Amyotrophic lateral sclerosis | Testosterone ↑ Neurosteroids ↑↓ Adrenal steroidogenesis ↑↓ | Target specificity, off-target effects |
| VDAC1 peptides | Testosterone deficiency | Testosterone ↑ | Development, adoption |

Note: Symbol ↑ denotes positive effect and symbol ↓ denotes negative effect.

Abbreviations: GnRH, gonadotropin releasing hormone; LH, luteinizing hormone; TSPO, translocator protein; VDAC1, voltage-dependent anion channel 1. Source: Ide et al. 16

However, this process, which is disrupted in compromised Leydig cells, causing disruptions to mitochondrial function, cellular homeostasis, and steroidogenesis. 58,65

3.3 | Reduced mitochondrial function of Leydig cells

Leydig cell steroidogenic function and cellular bioenergetics are integrally linked to one another, as steroidogenesis requires reliable mitochondrial membrane potential and ATP synthesis. 72,73 Mitochondrial dynamics such as fission, fusion, biogenesis, and mitophagy are, therefore, required for sustainable steroidogenic capacity. 41 The clearance of dysfunction mitochondria is mediated by PTEN induced kinase

1/parkin RBR E3 ubiquitin ligase (PINK1/PARKIN) interactions⁶⁹ and the generation of new mitochondria, mitochondrial biogenesis, is regulated by the genes nuclear respiratory factor 1/2 (*Nrf1/2*) and transcription factor A, mitochondrial (*Tfam*).⁷⁴ The trafficking of molecules across the mitochondrial membranes is mediated through a variety of mitochondrial contact sites, pores, and transporters all of which are regulated by mitofusion 1/2 (*Mfn1/2*), optic atrophy 1 (*Opa1*), and dynamin-related protein 1 (*Drp1*).⁷¹ Aging leads to a decline in these genes' expression systemically across many tissues,⁷⁵ and the reduction of steroidogenic capacity in aging Leydig cells in particular is driven by this mitochondrial dysfunction.⁵⁸ When compared with healthy cells, aged Leydig cells present depressed ATP levels, mitochondrial biogenesis, and mitophagy. Moreover, the expression of genes regulating these mitochondrial dynamics are decreased.⁵⁸



4 | ENDOGENOUS TARGETS FOR TESTOSTERONE RECOVERY THERAPY

The role of numerous SITE proteins and steroidogenic regulators have been investigated to identify endogenous therapeutic targets that induce steroid hormone formation. Several proteins within the cytosol and mitochondria mediate cholesterol translocation from intracellular stores to the OMM where the SITE complex resides.³⁸ Rone et al.³⁸ investigated the role of numerous steroidogenic and mitochondrial dynamic proteins to elucidate their role in steroidogenesis. Such investigations revealed that knocking down OPA1, VDAC1, and ATAD3A had no effect on membrane permeable steroid formation, However, VDAC1 and ATAD3A knockdowns did reduce hormone-induced steroidogenesis, suggesting that OPA1 is not critical for hormone-induced steroidogenesis.³⁸ Recently, it was shown that upregulating OPA1 via pharmacological and transfection methods increased TSPO expression and steroid hormone formation in both basal and hormone-stimulated dysfunctional Leydig cells, suggesting that OPA1 may play a role in the regulation and formation of the SITE complex.⁷⁶ Progress has been made in targeting SITE proteins to ameliorate testosterone decline. Studies on TSPO ligands and 14-3-3ε peptides (VDAC1 peptides) have offered potential therapeutic strategies for inducing endogenous testosterone formation. While TSPO ligands enhance cholesterol translocation, VDAC1 peptides are designed to block the negative regulation of steroidogenesis.34,51,77-80

4.1 | TSPO ligands

Engagement of the OMM protein TSPO via a drug ligand-induced activation stimulates steroid hormone production in vitro and in vivo in rats and mice. 48.61,77,78.81.82 TSPO possesses high affinity for cholesterol binding, which leads to its subsequent translocation to the IMM for side-chain cleavage by CYP11A1 producing pregnenolone. 83.84 TSPO's C terminus plays a key role in the uptake of cholesterol from the cytosol and translocation into the mitochondria, 85.86 and disruption of the protein within steroidogenic cells disrupts mitochondrial cholesterol transport and steroid formation. 84.87 Steroidogenesis and TSPO expression correlate with one another as shown by disruption of steroidogenesis with TSPO's age-related decline in vivo and its ablation in vitro. 62.87 Moreover, transfection of TSPO into TSPO-disrupted cells restores steroid formation, demonstrating its indispensable role in steroidogenesis. 87

Numerous studies have shown that drug ligands targeting TSPO produce enhanced steroid levels in both MA-10 tumorigenic Leydig cells and isolated primary Leydig cells, as well as increased serum testosterone levels. 77,78,81 However, serum LH levels may also become increased following TSPO drug ligand treatment likely because of an effect of the ligand on brain TSPO,88,89 suggesting that using this target may enhance testosterone biosynthesis by either stimulating the Leydig cell steroidogenic machinery and/or by elevating LH release.77 TSPO-specific ligands are also known to increase glucocorticoid and

corticosteroid levels⁶¹ and have been shown to affect neurosteroid production.^{90–92} Accordingly, the use of TSPO ligands as a therapeutic approach to treat neurological and psychiatric disorders have also been investigated.⁹³ Similarly, the use of TSPO ligands may also induce anxiolytic-like responses, as ligand treatment has been shown to counteract panic attacks in rodents.⁹⁴ While molecular entities targeting TSPO elevate serum testosterone levels, adrenal steroids and neurosteroids are also affected. Therefore, TSPO ligands have been proposed as therapeutic agents for the regulation of steroid hormones in the testis and brain. However, this lack of specificity remains an issue, as TSPO is expressed in numerous tissues.

4.2 | VDAC1 peptides

New insights into the role of $14-3-3\varepsilon$ in the regulation of steroidogenesis have made it a promising therapeutic target. 14-3-3 proteins regulate target proteins by altering activity, post-translational modifications, and subcellular localization. 95 LHR stimulation initiates the translocation of 14-3-3 ε to the OMM³³ and its recruitment to the TSPO-VDAC1 complex at Ser167 on VDAC1. There it competes with TSPO for VDAC1 binding and thus reduces cholesterol import.⁵¹ Blocking the interaction between 14-3-3ε and VDAC1 using cellpenetrating peptides induces steroid formation in vivo and ex vivo.80 Aghazadeh et al. 79 fused a component of the human immunodeficiency virus transcription factor 1 (TAT) with the predicted Ser167 binding motif on 14-3-3ε, creating a cell permeable VDAC1 peptide, TAT-VDAC1 containing Ser167 (TVS167), which competed with 14-3-3ε for VDAC1 binding. This reduced negative regulation of steroidogenesis by blocking the 14-3-3 ε binding to VDAC1, which led to increased steroidogenesis in vitro and in vivo. Given the homologous mechanisms of 14-3-3 ε between species, the TAT-based peptide offers a promising approach in humans. Although TAT peptides penetrate indiscriminately and 14-3-3 ε is found in numerous tissues, function is tissue specific.⁷⁹ TVS167 treatment did not significantly increase corticosterone levels in rats treated with the compound, demonstrating specificity to testicular Leydig cells.80 Additionally, TVS167 induced steroidogenesis independent of LH and would offer a major improvement in safety than TRT.96 The minimal bioactive sequence of the peptide was recently identified, and we ultimately generated bioactive stable peptide derivatives that can be administered orally and induce T formation in normal and hypogonadal animal models (manuscript in preparation). Moreover, they demonstrate safety, efficacy, and target specificity. 34,51,79,80 In summary, these first-inclass biologics make an excellent candidate for treatment of diseases caused by Leydig cell dysfunction over other pharmacologic or biologic strategies.

4.3 | Implantation of human Leydig-like cells

The generation of transplantable testosterone-producing cells offers another alternative for treating pathologies related to Leydig cell

dysfunction. Previously, it was shown that mesenchymal stem cells (MSCs) were able to differentiate into testosterone-producing Levdig cells, suggesting that healthy Leydig cell populations could be transplanted into hypogonadal patients.⁹⁷ However, MSC isolation produces limited cell numbers and reduces the clinical application of this method. Recent developments have revealed human Leydiglike cells (hLLCs) can be generated from human-induced pluripotent stem cells (hiPSCs), which are highly expandable in cell culture. 98,99 Li et al. 98 demonstrated that hLLCs producing steroidogenic gene expression, steroidogenic enzymes, and testosterone could be generated by differentiating early mesenchymal progenitors from hiPSCs while overexpressing steroidogenic factor 1 in culture with dibutyryl-cAMP, recombinant desert hedgehog, and human chorionic gonadotropin. Given their clinical viability, the implantation of hLLCs would represent a monumental step forward in treating diseases related to Leydig cell dysfunction. This strategy could restore testosterone levels by replenishing testosterone-producing cell populations in the testicular environment, leading to the production of endogenous testosterone formation.

5 | CONCLUSIONS AND FUTURE DIRECTIONS

Testosterone deficiency impacts the quality of life and wellbeing for millions of men worldwide, with only limited treatments having undesirable off-target effects. 16 New understanding of the molecular interactions producing testosterone has laid the foundation for the development of novel therapeutic strategies. Identification of the hormonally regulated multiprotein Steroidogenic InteracTomE complex³⁸ and the deeper understanding of hormonal stimulation and cholesterol translocation from cytosolic stores across the outer mitochondrial membrane and into the inner mitochondrial membrane for side-chain cleavage demonstrates numerous therapeutic targets for various indications related to hormone insufficiency (Table 1). 16,18 However, their effects on neurosteroids, adrenal steroids, and the hypothalamicpituitary-gonadal axis have remained a barrier to safe and efficacious treatment of testosterone deficiency. Apart from voltage-dependent anion channel 1 peptides, existing strategies have lacked specificity for testicular Leydig cells and, therefore, have raise concerns regarding off-target effects (Figure 2). Voltage-dependent anion channel 1 peptides are first-in-class biologics that offer a novel approach for rescuing intratesticular and serum testosterone formation in hormonally mediated diseases.^{79,80} These therapeutics could be used to restore endogenous testosterone formation and restore wellbeing for millions of aging men worldwide.

There are additional mechanisms to uncover. The movement of cholesterol between the mitochondrial membranes, the relationship between aging and the Leydig cell oxidative environment, and age-dependent protein-protein interactions remain elusive and are active areas of research.²⁴ With more information we may determine the cause of reduced testosterone and develop interventions that may maintain Leydig cell function. Moreover, targeting the molecular deteriorations that differ between aging Leydig cells and other

aging steroidogenic tissues could lead to additional testis-specific strategies.

AUTHOR CONTRIBUTIONS

The authors contributed equally as they conceptualized the content of this review, drafted the manuscript, and edited and reviewed the final manuscript.

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CONFLICTS OF INTEREST

Vassilios Papadopoulos is named inventor on patents filed with U.S.P.T.O. and other international agencies on therapeutics for the induction of endogenous steroidogenesis and licensed by McGill University to IASO BioMed. Vassilios Papadopoulos received stock and support for research from IASO BioMed when at the Research Institute of the McGill University Health Centre, Montreal, Canada.

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REFERENCES

- Zirkin BR, Papadopoulos V. Leydig cells: formation, function, and regulation. Biol Reprod. 2018;99(1):101-111.
- Cattabiani C, Basaria S, Ceda GP, et al. Relationship between testosterone deficiency and cardiovascular risk and mortality in adult men. J Endocrinol Invest. 2012;35(1):104-120.
- 3. Bhasin S, Brito JP, Cunningham GR, et al. Testosterone therapy in men with hypogonadism: an Endocrine Society Clinical Practice Guideline. *J Clin Endocrinol Metab*. 2018;103(5):1715-1744.
- Payne AH, Hardy MP. The Leydig Cell in Health and Disease. Springer Science & Business Media; 2007.
- Fraietta R, Zylberstejn DS, Esteves SC. Hypogonadotropic hypogonadism revisited. Clinics. 2013;68(suppl 1 (S1)):81-88.
- Kolettis PN, Purcell ML, Parker W, Poston T, Nangia AK. Medical testosterone: an iatrogenic cause of male infertility and a growing problem. *Urology*. 2015;85(5):1068-1073.
- 7. Reddy DS. Neurosteroids: endogenous role in the human brain and therapeutic potentials. *Prog Brain Res.* 2010;186:113-137.
- Li L, Papadopoulos V. Advances in stem cell research for the treatment of primary hypogonadism. Nat Rev Urol. 2021;18(8):487-507.
- Barbonetti A, D'Andrea S, Francavilla S. Testosterone replacement therapy. Andrology. 2020;8(6):1551-1566.
- Mulhall JP, Trost LW, Brannigan RE, et al. Evaluation and management of testosterone deficiency: AUA guideline. J Urol. 2018;200(2):423-432.
- McBride JA, Coward RM. Recovery of spermatogenesis following testosterone replacement therapy or anabolic-androgenic steroid use. *Asian J Androl.* 2016;18(3):373-380.
- Westfield G, Kaiser UB, Lamb DJ, Ramasamy R. Short-acting testosterone: more physiologic? Front Endocrinol. 2020;11:572465.
- Basaria S, Coviello AD, Travison TG, et al. Adverse events associated with testosterone administration. N Engl J Med. 2010;363(2): 109-122.
- Finkle WD, Greenland S, Ridgeway GK, et al. Increased risk of non-fatal myocardial infarction following testosterone therapy prescription in men. PLoS One. 2014;9(1):e85805.

- Vigen R, O'Donnell CI, Baron AE, et al. Association of testosterone therapy with mortality, myocardial infarction, and stroke in men with low testosterone levels. JAMA. 2013;310(17):1829-1836.
- Ide V, Vanderschueren D, Antonio L. Treatment of men with central hypogonadism: alternatives for testosterone replacement therapy. *Int* J Mol Sci. 2020;22(1):21.
- Wang C, Swerdloff RS. Should the nonaromatizable androgen dihydrotestosterone be considered as an alternative to testosterone in the treatment of the andropause? J Clin Endocrinol Metab. 2002;87(4):1462-1466.
- Aydogdu A, Swerdloff RS. Emerging medication for the treatment of male hypogonadism. Expert Opin Emerg Drugs. 2016;21(3):255-266.
- Maggi M, Buvat J, Corona G, Guay A, Torres LO. Hormonal causes of male sexual dysfunctions and their management (hyperprolactinemia, thyroid disorders, GH disorders, and DHEA). J Sex Med. 2013;10(3):661-677.
- Loves S, Ruinemans-Koerts J, de Boer H. Letrozole once a week normalizes serum testosterone in obesity-related male hypogonadism. *Eur J Endocrinol*. 2008;158(5):741-747.
- Hero M, Toiviainen-Salo S, Wickman S, Makitie O, Dunkel L. Vertebral morphology in aromatase inhibitor-treated males with idiopathic short stature or constitutional delay of puberty. *J Bone Miner Res.* 2010:25(7):1536-1543.
- Clavijo RI, Hsiao W. Update on male reproductive endocrinology. *Transl Androl Urol.* 2018;7(suppl 3):S367-S372.
- Rone MB, Fan J, Papadopoulos V. Cholesterol transport in steroid biosynthesis: role of protein-protein interactions and implications in disease states. *Biochim Biophys Acta*. 2009;1791(7):646-658.
- 24. Papadopoulos V, Zirkin BR. Leydig cell aging: molecular mechanisms and treatments. *Vitam Horm.* 2021;115:585-609.
- 25. Besman MJ, Yanagibashi K, Lee TD, Kawamura M, Hall PF, Shively JE. Identification of des-(Gly-Ile)-endozepine as an effector of corticotropin-dependent adrenal steroidogenesis: stimulation of cholesterol delivery is mediated by the peripheral benzodiazepine receptor. Proc Natl Acad Sci U S A. 1989;86(13):4897-4901.
- Boujrad N, Hudson JR, Jr., Papadopoulos V. Inhibition of hormonestimulated steroidogenesis in cultured Leydig tumor cells by a cholesterol-linked phosphorothioate oligodeoxynucleotide antisense to diazepam-binding inhibitor. *Proc Natl Acad Sci U S A*. 1993;90(12):5728-5731.
- Schultz R, Pelto-Huikko M, Alho H. Expression of diazepam binding inhibitor-like immunoreactivity in rat testis is dependent on pituitary hormones. *Endocrinology*. 1992;130(6):3200-3206.
- 28. Bose HS, Lingappa VR, Miller WL. Rapid regulation of steroidogenesis by mitochondrial protein import. *Nature*. 2002;417(6884):87-91.
- Clark BJ, Wells J, King SR, Stocco DM. The purification, cloning, and expression of a novel luteinizing hormone-induced mitochondrial protein in MA-10 mouse Leydig tumor cells. Characterization of the steroidogenic acute regulatory protein (StAR). J Biol Chem. 1994;269(45):28314-28322.
- Miller WL. Steroidogenic acute regulatory protein (StAR), a novel mitochondrial cholesterol transporter. Biochim Biophys Acta. 2007;1771(6):663-676.
- 31. Arakane F, Sugawara T, Nishino H, et al. Steroidogenic acute regulatory protein (StAR) retains activity in the absence of its mitochondrial import sequence: implications for the mechanism of StAR action. *Proc Natl Acad Sci U S A.* 1996;93(24):13731-13736.
- Baker BY, Epand RF, Epand RM, Miller WL. Cholesterol binding does not predict activity of the steroidogenic acute regulatory protein, StAR. J Biol Chem. 2007;282(14):10223-10232.
- Aghazadeh Y, Rone M, Blonder J, et al. Hormone-induced 14-3-3γ adaptor protein regulates steroidogenic acute regulatory protein activity and steroid biosynthesis in MA-10 Leydig cells. *J Biol Chem*. 2012;287(19):15380-15394.

- Aghazadeh Y, Papadopoulos V. The role of the 14-3-3 protein family in health, disease, and drug development. *Drug Discov Today*. 2016:21(2):278-287.
- Aghazadeh Y, Venugopal S, Martinez-Arguelles DB, Boisvert A, Blonder J, Papadopoulos V. Identification of Sec23ip, Part of 14-3-3gamma protein network, as a regulator of acute steroidogenesis in MA-10 leydig cells. *Endocrinology*. 2020;161(2):bqz036.
- Liu J, Rone MB, Papadopoulos V. Protein–protein interactions mediate mitochondrial cholesterol transport and steroid biosynthesis. *J Biol Chem.* 2006;281(50):38879-38893.
- Issop L, Rone MB, Papadopoulos V. Organelle plasticity and interactions in cholesterol transport and steroid biosynthesis. Mol Cell Endocrinol. 2013;371(1-2):34-46.
- Rone MB, Midzak AS, Issop L, et al. Identification of a dynamic mitochondrial protein complex driving cholesterol import, trafficking, and metabolism to steroid hormones. *Mol Endocrinol*. 2012;26(11):1868-1882.
- Issop L, Fan J, Lee S, et al. Mitochondria-associated membrane formation in hormone-stimulated Leydig cell steroidogenesis: role of ATAD3. Endocrinology. 2015;156(1):334-345.
- Crompton M, Barksby E, Johnson N, Capano M. Mitochondrial intermembrane junctional complexes and their involvement in cell death. Biochimie. 2002;84(2-3):143-152.
- 41. Duarte A, Poderoso C, Cooke M, et al. Mitochondrial fusion is essential for steroid biosynthesis. *PloS One*. 2012;7(9):e45829.
- Hauet T, Yao ZX, Bose HS, et al. Peripheral-type benzodiazepine receptor-mediated action of steroidogenic acute regulatory protein on cholesterol entry into leydig cell mitochondria. *Mol Endocrinol*. 2005:19(2):540-554.
- Fan J, Liu J, Culty M, Papadopoulos V. Acyl-coenzyme A binding domain containing 3 (ACBD3; PAP7; GCP60): an emerging signaling molecule. *Prog Lipid Res.* 2010;49(3):218-234.
- 44. Dyson MT, Jones JK, Kowalewski MP, et al. Mitochondrial A-kinase anchoring protein 121 binds type II protein kinase A and enhances steroidogenic acute regulatory protein-mediated steroidogenesis in MA-10 mouse Leydig tumor cells. *Biol Reprod.* 2008;78(2): 267-277.
- Delavoie F, Li H, Hardwick M, et al. In vivo and in vitro peripheral-type benzodiazepine receptor polymerization: functional significance in drug ligand and cholesterol binding. *Biochemistry*. 2003;42(15):4506-4519.
- Krueger KE, Papadopoulos V. Peripheral-type benzodiazepine receptors mediate translocation of cholesterol from outer to inner mitochondrial membranes in adrenocortical cells. J Biol Chem. 1990;265(25):15015-15022.
- Yanagibashi K, Ohno Y, Nakamichi N, et al. Peripheral-type benzodiazepine receptors are involved in the regulation of cholesterol side chain cleavage in adrenocortical mitochondria. *J Biochem*. 1989;106(6):1026-1029.
- Papadopoulos V, Fan J, Zirkin B. Translocator protein (18 kDa): an update on its function in steroidogenesis. *J Neuroendocrinol*. 2018;30(2):e12500.
- McEnery MW, Snowman AM, Trifiletti RR, Snyder SH. Isolation of the mitochondrial benzodiazepine receptor: association with the voltagedependent anion channel and the adenine nucleotide carrier. *Proc Natl Acad Sci U S A.* 1992;89(8):3170-3174.
- Tremblay JJ. Molecular regulation of steroidogenesis in endocrine Leydig cells. Steroids. 2015;103:3-10.
- Aghazadeh Y, Ye X, Blonder J, Papadopoulos V. Protein modifications regulate the role of 14-3-3gamma adaptor protein in cAMP-induced steroidogenesis in MA-10 Leydig cells. J Biol Chem. 2014;289(38):26542-26553.
- 52. Martin LJ, Tremblay JJ. Nuclear receptors in Leydig cell gene expression and function. *Biol Reprod*. 2010;83(1):3-14.

- Gorostizaga A, Cornejo Maciel F, Brion L, Maloberti P, Podesta EJ, Paz C. Tyrosine phosphatases in steroidogenic cells: regulation and function. Mol Cell Endocrinol. 2007;265-266:131-137.
- Epstein LF, Orme-Johnson NR. Acute action of luteinizing hormone on mouse Leydig cells: accumulation of mitochondrial phosphoproteins and stimulation of testosterone synthesis. Mol Cell Endocrinol. 1991;81(1-3):113-126.
- Luo L, Chen H, Stocco DM, Zirkin BR. Leydig cell protein synthesis and steroidogenesis in response to acute stimulation by luteinizing hormone in rats. *Biol Reprod.* 1998;59(2):263-270.
- Bose HS, Sugawara T, Strauss JF, 3rd, Miller WL, International Congenital Lipoid Adrenal Hyperplasia Consortium. The pathophysiology and genetics of congenital lipoid adrenal hyperplasia. N Engl J Med. 1996;335(25):1870-1878.
- 57. Galano M, Li Y, Li L, Sottas C, Papadopoulos V. Role of constitutive STAR in Leydig cells. *Int J Mol Sci.* 2021;22(4):2021.
- Sokanovic SJ, Baburski AZ, Kojic Z, Medar MLJ, Andric SA, Kostic TS.
 Aging-related increase of cGMP disrupts mitochondrial homeostasis in leydig cells. J Gerontol A Biol Sci Med Sci. 2021;76(2):177-186.
- Owen DR, Fan J, Campioli E, et al. TSPO mutations in rats and a human polymorphism impair the rate of steroid synthesis. *Biochem J*. 2017;474(23):3985-3999.
- Fan J, Campioli E, Midzak A, Culty M, Papadopoulos V. Conditional steroidogenic cell-targeted deletion of TSPO unveils a crucial role in viability and hormone-dependent steroid formation. *Proc Natl Acad Sci* U S A. 2015;112(23):7261-7266.
- Papadopoulos V, Aghazadeh Y, Fan J, Campioli E, Zirkin B, Midzak A. Translocator protein-mediated pharmacology of cholesterol transport and steroidogenesis. Mol Cell Endocrinol. 2015;408:90-98.
- Culty M, Luo L, Yao ZX, Chen H, Papadopoulos V, Zirkin BR. Cholesterol transport, peripheral benzodiazepine receptor, and steroidogenesis in aging Leydig cells. *J Androl*. 2002;23(3):439-447.
- 63. Beattie MC, Adekola L, Papadopoulos V, Chen H, Zirkin BR. Leydig cell aging and hypogonadism. *Exp Gerontol.* 2015;68:87-91.
- Jendrach M, Mai S, Pohl S, Voth M, Bereiter-Hahn J. Short- and longterm alterations of mitochondrial morphology, dynamics and mtDNA after transient oxidative stress. *Mitochondrion*. 2008;8(4):293-304.
- 65. Jang JY, Blum A, Liu J, Finkel T. The role of mitochondria in aging. *J Clin Invest*. 2018;128(9):3662-3670.
- 66. Chen H, Cangello D, Benson S, et al. Age-related increase in mitochondrial superoxide generation in the testosterone-producing cells of Brown Norway rat testes: relationship to reduced steroidogenic function? *Exp Gerontol.* 2001;36(8):1361-1373.
- 67. Hornsby PJ. Steroid and xenobiotic effects on the adrenal cortex: mediation by oxidative and other mechanisms. *Free Radic Biol Med.* 1989;6(1):103-115.
- Peltola V, Huhtaniemi I, Metsa-Ketela T, Ahotupa M. Induction of lipid peroxidation during steroidogenesis in the rat testis. *Endocrinology*. 1996;137(1):105-112.
- Palikaras K, Lionaki E, Tavernarakis N. Mechanisms of mitophagy in cellular homeostasis, physiology and pathology. *Nat Cell Biol*. 2018;20(9):1013-1022.
- 70. Kauppila TES, Kauppila JHK, Larsson NG. Mammalian mitochondria and aging: an update. *Cell Metab*. 2017;25(1):57-71.
- Friedman JR, Nunnari J. Mitochondrial form and function. *Nature*. 2014;505(7483):335-343.
- Midzak AS, Chen H, Aon MA, Papadopoulos V, Zirkin BR. ATP synthesis, mitochondrial function, and steroid biosynthesis in rodent primary and tumor Leydig cells. *Biol Reprod*. 2011;84(5):976-985.
- Allen JA, Shankara T, Janus P, et al. Energized, polarized, and actively respiring mitochondria are required for acute Leydig cell steroidogenesis. *Endocrinology*. 2006;147(8):3924-3935.
- 74. Li PA, Hou X, Hao S. Mitochondrial biogenesis in neurodegeneration. *J Neurosci Res.* 2017;95(10):2025-2029.

- Moreira OC, Estebanez B, Martinez-Florez S, de Paz JA, Cuevas MJ, Gonzalez-Gallego J. Mitochondrial function and mitophagy in the elderly: effects of exercise. Oxid Med Cell Longev. 2017;2017:2012798.
- Garza S, Galano M, Li L, Papadopoulos V. Role of TSPO in mitochondrial fusion and function in MA-10 mouse tumor Leydig cells. Andrology. 2022;10:89.
- Chen F, Lu H, Chen P, et al. Acute effects of the translocator protein drug ligand FGIN-1-27 on serum testosterone and luteinizing hormone levels in male Sprague-Dawley ratsdagger. *Biol Reprod.* 2019;100(3):824-832.
- Chung JY, Chen H, Midzak A, Burnett AL, Papadopoulos V, Zirkin BR. Drug ligand-induced activation of translocator protein (TSPO) stimulates steroid production by aged brown Norway rat Leydig cells. *Endocrinology*, 2013;154(6):2156-2165.
- Aghazadeh Y, Martinez-Arguelles DB, Fan J, Culty M, Papadopoulos V. Induction of androgen formation in the male by a TAT-VDAC1 fusion peptide blocking 14-3-3varepsilon protein adaptor and mitochondrial VDAC1 interactions. Mol Ther. 2014;22(10):1779-1791.
- Papadopoulos V. Testosterone recovery therapy targeting dysfunctional Leydig cells. In: North American Testis Workshop. May 4-7, La Jolla, CA, USA. 2022.
- Musicki B, Karakus S, La Favor JD, et al. TSPO ligand FGIN-1-27 controls priapism in sickle cell mice via endogenous testosterone production. J Cell Physiol. 2021;236(4):3073-3082.
- Papadopoulos V. Peripheral-type benzodiazepine/diazepam binding inhibitor receptor: biological role in steroidogenic cell function. *Endocr Rev.* 1993;14(2):222-240.
- 83. Scarf AM, Kassiou M. The translocator protein. *J Nucl Med*. 2011;52(5):677-680.
- Midzak A, Rone M, Aghazadeh Y, Culty M, Papadopoulos V. Mitochondrial protein import and the genesis of steroidogenic mitochondria. Mol Cell Endocrinol. 2011;336(1-2):70-79.
- Li H, Papadopoulos V. Peripheral-type benzodiazepine receptor function in cholesterol transport. Identification of a putative cholesterol recognition/interaction amino acid sequence and consensus pattern. *Endocrinology*. 1998;139(12):4991-4997.
- 86. Li H, Yao Z, Degenhardt B, Teper G, Papadopoulos V. Cholesterol binding at the cholesterol recognition/interaction amino acid consensus (CRAC) of the peripheral-type benzodiazepine receptor and inhibition of steroidogenesis by an HIV TAT-CRAC peptide. Proc Natl Acad Sci U S A. 2001;98(3):1267-1272.
- Papadopoulos V, Amri H, Li H, Boujrad N, Vidic B, Garnier M. Targeted disruption of the peripheral-type benzodiazepine receptor gene inhibits steroidogenesis in the R2C Leydig tumor cell line. *J Biol Chem.* 1997;272(51):32129-32135.
- Wang HJ, Fan J, Papadopoulos V. Translocator protein (Tspo) gene promoter-driven green fluorescent protein synthesis in transgenic mice: an in vivo model to study Tspo transcription. *Cell Tissue Res.* 2012;350(2):261-275.
- 89. Nutma E, Ceyzeriat K, Amor S, et al. Cellular sources of TSPO expression in healthy and diseased brain. *Eur J Nucl Med Mol Imaging*. 2021;49(1):146-163.
- Lin Y, Cheung G, Espinoza N, Papadopoulos V. Function, regulation, and pharmacological effects of pregnenolone in the central nervous system. Curr Opin Endocr Metab Res. 2022;22:100310.
- Rupprecht R, Papadopoulos V, Rammes G, et al. Translocator protein (18 kDa)(TSPO) as a therapeutic target for neurological and psychiatric disorders. Nat Rev Drug Discov. 2010;9(12):971-988.
- 92. Lin Y, Cheung G, Porter E, Papadopoulos V. The neurosteroid pregnenolone is synthesized by a mitochondrial P450 enzyme other than CYP11A1 in human glial cells. *J Biol Chem.* 2022;298(7):102110.
- 93. Rupprecht R, Papadopoulos V, Rammes G, et al. Translocator protein (18 kDa) (TSPO) as a therapeutic target for neurological and psychiatric disorders. *Nat Rev Drug Discov*. 2010;9(12):971-988.

- 94. Rupprecht R, Rammes G, Eser D, et al. Translocator protein (18 kD) as target for anxiolytics without benzodiazepine-like side effects. *Science*. 2009;325(5939):490-493.
- 95. Tzivion G, Shen YH, Zhu J. 14-3-3 proteins; bringing new definitions to scaffolding. *Oncogene*. 2001;20(44):6331-6338.
- 96. Campanella M. Peptide targeting of mitochondria elicits testosterone formation. *Mol Ther.* 2014;22(10):1727-1729.
- 97. Yazawa T, Mizutani T, Yamada K, et al. Differentiation of adult stem cells derived from bone marrow stroma into Leydig or adrenocortical cells. *Endocrinology*. 2006;147(9):4104-4111.
- 98. Li L, Li Y, Sottas C, et al. Directing differentiation of human induced pluripotent stem cells toward androgen-producing Leydig cells rather

- than adrenal cells. Proc Natl Acad Sci U S A. 2019;116(46):23274-
- 99. Yamanaka S. Induced pluripotent stem cells: past, present, and future. Cell Stem Cell. 2012;10(6):678-684.

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